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			TE	EST RE				
Reg. No.	: 104100649	Reg. Date	:04-Apr-20	21 12:01	Ref.No :	Collected On	: 04-Apr-2021 12:01	
Name	: DIGVIJAY SINGH RATHORE MR					Reported Date	Reported Date : 04-Apr-2021	
Age	: 23 Years	Gender	: Male	Pass.	No. :	Dispatch At	:	
Ref. By	:					Tele No.	:	
Location	: SURENDRANAGAR COVID COLLECTION CENTER @ SURENDRANAGAR							

Test Name	Results	Units	Bio. Ref. Interval							
MOLECULAR A	NALYSIS FOR QUALITATIV	E DETECTION	OF SARS-CoV-2.							
<u>Type of sample : Nasopharyngeal swab and Oropharyngeal swab.</u>										
Methodo	logy : Real time PCR.	ICMR NO	:UNIPA001							
ORF 1ab	Negative									
N Gene	Negative									
S Gene	Negative									
MS2 GENE(Internal Control)	Pass									
Interpretation										
2019-nCoV	NEGATIVE									

Note :

For results with S-Gene negative and other two genes positive: These samples may have 69-70del S gene mutation which is usually associated with, but not limited to B.1.1.7 variant (UK VOC-202012/01). Since our assay is designed to detect multiple genetic targets (ORF and N genes along with S gene), the overall test sensitivity is not impacted by this variant. (Ref: <u>https://www.fda.gov/medical-devices/letters-health-care-providers/genetic-variants-sars-cov-2-may-lead-false-negative-results-molecular-tests-detection-sars-cov-2)</u>

1 - Test report should be correlated with the clinical presentation and findings.

2 - The LOD for the three target genes is 10 copies/reaction.

3 - A negative result does not rule out 2019-nCoV and should not be used as the sole basis for treatment or other patient management decisions.

4 - A number of factors could lead to a negative result in an infected individual including 1) Poor quality of the specimen, containing inadequate patient material or non-representative specimen 2) The specimen was collected late or very early in the infection. Optimum specimen types and timing for peak viral levels during infections caused by 2019-nCoV have not been determined. Collection of multiple samples from the same patient may be necessary to detect the virus 3) The specimen was not handled and shipped appropriately 4) Technical reasons inherent in the test (like Virus mutation or PCR inhibition) 5) Inadequate numbers of organisms are present in the specimen

5 - Reports will be provided to the treating physician, who is requested to communicate the same to the patient and follow MOHFW policy for isolation, quarantine and treatment of all positive cases along with contact tracing as recommended.

6 - Repeat sampling and testing of lower respiratory specimen is strongly recommended in severe or progressive disease. 7 -The repeat specimens may be considered after a gap of 2-4 days after the collection of the first specimen for additional testing if required.

8 - Categories of viral load is based on Cycle threshold (Ct) detected by RT PCR.

9 - High viral load: up to 23; Moderate viral load: 24 to 31; Low/Mild viral load: 32 to 35

End Of Report

Test done from collected sample.

This is an electronically authenticated report.



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